

Methane dynamics in an estuarine brackish *Cyperus malaccensis* marsh: Production and porewater concentration in soils, and net emissions to the atmosphere over five years

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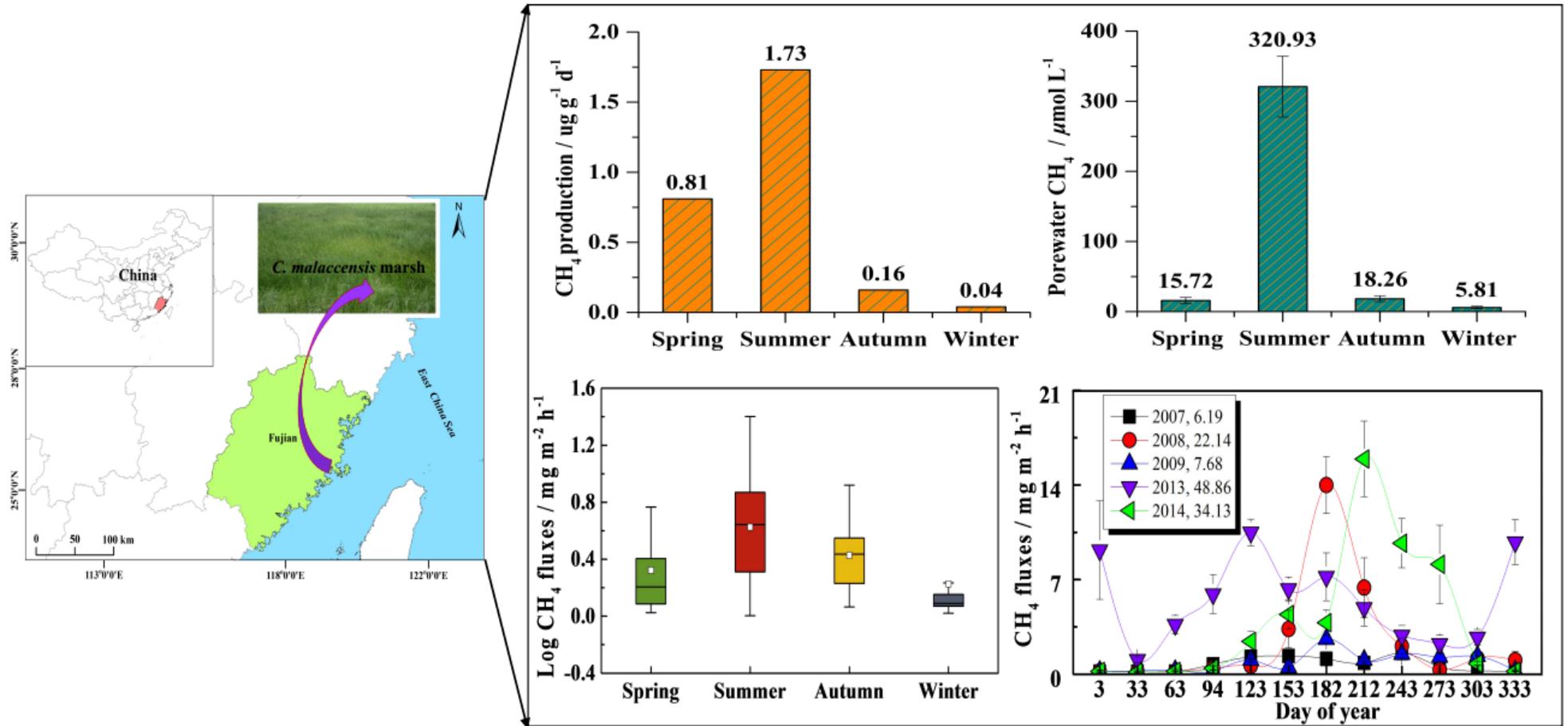
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HIGHLIGHTS

- Coastal marsh CH₄ emissions had strong inter-annual and seasonal variability.
- High CH₄ emissions were accompanied by high CH₄ production and porewater CH₄ concentrations.
- Brackish marshes in subtropical estuaries were important sources of CH₄.
- Temperature, precipitation and salinity were the best predictors of CH₄ emissions

Graphical Abstract



**Methane dynamics in an estuarine brackish *Cyperus malaccensis* marsh:
Production and porewater concentration in soils, and net emissions to the
atmosphere over five years**

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ABSTRACT

Wetlands can potentially affect global climate change through their role in modulating the atmospheric concentrations of methane (CH₄). Their overall CH₄ emissions, however, remain the greatest uncertainty in the global CH₄ budget. One reason for this is the paucity of long-term field measurements to characterize the variability of CH₄ emissions from different types of wetlands. In this study, we quantified CH₄ emissions from a brackish, oligohaline *Cyperus malaccensis* marsh ecosystem in the Min River Estuary in southeast China over five years. Our results showed substantial temporal variability of CH₄ emissions from this brackish marsh, with hourly fluxes ranging from 0.7±0.6 to 5.1±3.7 mg m⁻² h⁻¹ (mean ± 1 SD) during the study period. The inter-annual variability of CH₄ emissions was significantly correlated with changes in soil temperature, precipitation and salinity, which highlighted the importance of long-term observations in understanding wetland CH₄ dynamics. Distinct seasonal patterns in soil CH₄ production rates and porewater CH₄ concentrations also were observed, and were both positively correlated with CH₄ emissions. The seasonal variations of CH₄ emissions and production were highly correlated with salinity and porewater sulfate levels. The mean annual CH₄ efflux from our site over the five-year period was 23.8±18.1 g CH₄ m⁻² yr⁻¹, indicating that subtropical brackish tidal marsh ecosystems could release a large amount of CH₄ into the atmosphere. Our findings further highlight the need to obtain high-frequency and continuous field measurements over the long term at multiple spatial scales to improve our current estimates of wetland CH₄ emissions.

Keywords: Methane; Net emissions; Soil production; Porewater; Temporal variation; Estuarine marsh

1. Introduction

The increasing worldwide concern over global climate change and its effects on environmental and human well-beings calls for a better understanding of the magnitude of global greenhouse gas emissions ([Tong et al., 2010](#)). Methane (CH₄) is a potent greenhouse gas with a global warming potential 34 times higher than that of CO₂ per mass unit over a 100-year time scale, and contributes to approximately 20% of the global radiative forcing ([IPCC, 2013](#)). Global atmospheric CH₄ levels have increased by threefold since 1750, reaching 1845±2 ppb in 2015 ([World Meteorological Organization, 2016](#)). Quantifying the potential source strength of various ecosystems has become one of the top priorities for improving the future predictions of CH₄ emissions.

Wetlands are estimated to contribute 20–39% of the global CH₄ emissions ([Laanbroek, 2010](#)), with natural wetlands being the single largest source of CH₄. Over the past few decades, considerable efforts were made to quantify CH₄ emissions from different natural wetlands around the world (e.g. [Bubier et al., 1994](#); [Kutzbach et al., 2004](#); [Hendriks et al., 2010](#); [Tong et al., 2012](#)). However, the majority of these field campaigns were carried out over a relatively short period of not more than two years, which provided little knowledge of the inter-annual variability of CH₄ emissions from most types of wetlands other than a few exceptions in northern wetlands, e.g. [Song et al. \(2009\)](#), [Jackowicz-Korczyński et al. \(2010\)](#), and [Moore et al. \(2011\)](#). Long-term observations over multiple seasons and years are critical for determining accurate ecosystem CH₄ budgets ([Song et al., 2009](#)). In addition, the availability of long-term data set will improve ecosystem modelling by providing inputs for model calibration and validation, as well as insights on the key factors regulating wetland CH₄ emissions into the atmosphere ([Tian et al., 2008](#); [Song](#)

et al., 2009).

Coastal wetlands, located at the interface between the terrestrial and marine environments, are biogeochemically important ecosystems that span widely from the arctic to the tropical zones (Chmura et al., 2003; Wang et al., 2016). Previous studies have shown that the sediments in coastal wetlands are generally small atmospheric sources (Bartlett & Harriss, 1993; Poffenbarger et al., 2011; Livesley & Andrusiak, 2012; Koebisch et al., 2013), or even weak sinks of CH₄ (Sun et al., 2013). The low CH₄ source strength of coastal wetlands is mainly because of the relatively high sulfate concentrations in marine waters, which favour the activities of sulfate-reducing bacteria while at the same time hamper the metabolism of methanogens through intense competition for substrates (Poffenbarger et al., 2011; Callaway et al., 2012; Vizza et al., 2017). However, some short-term field studies provide evidence that large CH₄ emissions from wetlands can occur even when sulfate reduction is a dominant process (Lee et al., 2008; Marín-Muñiz et al., 2015; Holm Jr. et al., 2016). The high uncertainty associated with the magnitude and control of CH₄ emissions from coastal wetlands could partly be related to the inherently dynamic environment which introduces a large temporal variability of CH₄ fluxes that is not adequately accounted for by infrequent field measurements.

In this study, monthly CH₄ flux measurements were made in a subtropical tidal *Cyperus malaccensis* (shichito matgrass) marsh in the Min River Estuary in southeast China over five years between 2007-2009, and 2013-2014. We hypothesized that there would be significant seasonal and inter-annual variability in CH₄ emissions, which implies that flux estimates would be sensitive to the sampling frequency and study duration. We also investigated the temporal correlations between several environmental variables and soil CH₄ production rate, porewater

CH₄ concentration, and net CH₄ emissions.

2. Materials and methods

2.1. Site description

This study was carried out in the Shanyutan wetland (26°00'36"–26°03'42" N, 119°34'12"–119°40'40" E), the largest tidal wetland area (ca. 3120 ha) in the Min River Estuary, southeast China (Fig. 1). The Shanyutan wetland is influenced by a subtropical monsoonal climate, with a mean annual temperature of 19.6 °C and an annual precipitation of 1350 mm (Tong et al., 2010). The dominant vegetation species in the Shanyutan wetland included the native *Cyperus malaccensis* and *Phragmites australis*, as well as the invasive *Spartina alterniflora* (smooth cordgrass). The average height of *C. malaccensis* at the site was about 1.4 m. The study site was characterized by semi-diurnal tides, such that the soil surface was submerged for approximately 7 h over a 24 h cycle, and at other times, the soil surface was exposed to air (Tong et al., 2010). The average salinity of the tidal water was 4.2±2.5‰ (Tong et al., 2010).

2.2. Gas sampling and CH₄ flux estimation

Net CH₄ emissions were measured in the intertidal zone in the mid-western part of the Shanyutan wetland (26°01'46" N, 119°37'31" E), which was dominated by *C. malaccensis*, a widespread plant species at the site. Triplicate 1 m x 1 m plots, with a distance of < 5 m between plots, were established for regular measurement of CH₄ emissions in the *C. malaccensis* stand. CH₄ flux measurements were carried out monthly from early January to early December in 2007–2009 and 2013–2014. A wooden boardwalk was built to facilitate access to the study plots and minimize potential plot disturbance caused by field measurements. The wooden boardwalk and the study plots were damaged during a major typhoon event in 2010, thus we built a new

boardwalk and established new plots adjacent to the damaged ones (< 15 m apart) in 2012. During 2013–2014, we continued with gas flux measurements at the new plots.

CH₄ flux measurements were made using static closed chambers and gas chromatography techniques (Hirota et al., 2004; Song et al., 2009; Moore et al., 2011; Marín-Muñiz et al., 2015) with gas samples collected during the neap tides in the morning. The static chamber consisted of two parts: a 30 cm tall stainless steel bottom collar (length and width of 50 × 50 cm in 2007-2009, and 35 × 35 cm in 2013-2014) and a polyvinyl chloride top chamber (length, width and height of 50 × 50 × 170 cm in 2007-2009, and 35 × 35 × 140 cm in 2013-2014). The bottom collar was inserted into the marsh soils, leaving only 2 cm above the soil surface, approximately 10 days prior to the first flux measurement, and was then left in place for the duration of the study. A fan was installed inside the chamber to mix the headspace air during gas sampling. During each flux measurement, headspace air samples were drawn into air sampling bags (Dalian Delin Gas Packing Co., Ltd., China) at 10-minute intervals over a total duration of 30 min in each sampling plot. The total number of gas samples collected per year was 144 (12 months × 4 time intervals × 3 sites). CH₄ concentrations in the gas samples were determined using a gas chromatograph (GC-2010, Shimadzu, Kyoto, Japan) equipped with a flame ionization detector (FID). The rate of CH₄ emission (mg m⁻² h⁻¹) was calculated based on the slope of the linear regression between CH₄ concentration in the chamber headspace and time. The annual (cumulative) CH₄ emissions (*AE*, g CH₄ m⁻²) (Song et al., 2009; Moore et al., 2011; Xiang et al., 2015) were calculated using Eq. (1):

$$AE = \sum MF_i \times D_i \times 24 \quad (1)$$

where *MF_i* is the CH₄ flux at the *i*th month of the year (mg CH₄ m⁻² h⁻¹), and *D_i* is the number of

days in the i th month of the year.

2.3. Measurement of soil CH₄ production rate

Soil CH₄ production in coastal wetlands has distinct spatio-temporal heterogeneity that could be related to variations in thermal conditions and other abiotic factors (e.g. soil moisture, soil substrate, etc.) (Segers, 1998; Vizza et al., 2017). To assess the variability of soil CH₄ production rates across different depths in our marsh, triplicate sediment cores were randomly collected down to a depth of 100 cm in January (winter), March (spring), July (summer), and October (autumn) of 2012. Intact soil cores were collected using a steel sediment sampler (i.d. = 5 cm), sub-divided into ten sections at 10 cm intervals in the field, and then kept on ice in coolers and transported to the laboratory within 6 h. The rate of soil CH₄ production was measured following the method of Wachinger et al. (2000). The chambers (5 cm inner diameter, 12 cm height) used for the anoxic incubation of soil cores were made of polyoxymethylene, which was gas-impermeable and inert to CH₄. Before the start of incubation, the chambers were flushed with N₂ gas for 15 min to create an anaerobic condition (Wassmann et al., 1998). The cores were then incubated at *in situ* temperatures, i.e. 10.2, 17.5, 27.5, and 21.5 °C for winter, spring, summer, and autumn, respectively, for a duration of 15 days. We collected 5 mL gas samples from the chamber using a syringe at three day intervals ($n = 5$) over the course of the incubation, with N₂ gas being added after each gas sampling to re-establish the ambient atmospheric pressure. The CH₄ concentrations in gas samples were analysed immediately by gas chromatograph. The CH₄ production rates ($\mu\text{g CH}_4 \text{ g}^{-1}$ (dry weight) day^{-1}) were calculated based on the rate of change in chamber headspace CH₄ concentrations over a 3-day period (Wassmann et al., 1998). The total number of incubations made over the study period was 120 (3 replicates \times 4 seasons \times 10

depths).

2.4. Porewater collection and analysis of dissolved CH_4 and SO_4^{2-} concentrations

Porewater was sampled using the method of *in situ* dialysis (Ding et al., 2003; Ding et al., 2004a). A series of porewater tubes (5 cm inner diameter) (Ding et al., 2003), with sampling depths of 0–5, 5–10, 10–15, 15–20 and 20–25 cm, were permanently installed adjacent to each CH_4 flux measurement plot, leaving a 5-cm protrusion above the soil surface. The top of each tube was sealed tightly with a cover. Porewater samples were collected in triplicate at each depth interval in January (winter), March (spring), July (summer), and October (autumn) of 2012 and 2013. During each sampling campaign, approximately 10 mL of soil porewater was extracted using a syringe and discarded. Another 10 mL of porewater was then collected and transferred into a 20 mL pre-evacuated vial that was filled with 10 mL of pure N_2 gas (Xiang et al., 2015). About 0.2 mL of HgCl_2 solution was further injected into the porewater samples to inhibit bacterial activities without affecting the solubility of CH_4 in water (Butler and Elkins, 1991). The porewater samples were stored at about 4 °C in a cooler and transported immediately to the laboratory within 24 h for analysis. The sample vials were shaken vigorously for 10 min to establish an equilibrium in CH_4 concentrations between the dissolved phase in porewater and the gaseous phase in headspace. The headspace CH_4 concentrations were determined by gas chromatograph, and the dissolved CH_4 concentrations ($\mu\text{mol CH}_4 \text{ L}^{-1}$) in porewater were then calculated following the methods of Johnson et al. (1990) and Zhang et al. (2010).

To determine porewater SO_4^{2-} concentrations across different soil depths, another triplicate soil cores were collected down to a depth of 100 cm were collected in January (winter), March (spring), July (summer) and October (autumn) of 2012. The cores were split into ten sub-samples

at 10 cm intervals, which were then immediately sealed in a valve bag, kept on ice in coolers, and transported to the laboratory within 6 h. Upon return to the laboratory, porewater was extracted from the soils at each depth interval by centrifugation at 5000 rpm for 10 min (Cence® L550). The porewater samples were filtered with 0.45 µm acetate fibre membranes, and the SO₄²⁻ concentrations were determined using the barium chromate colorimetric method. The soil SO₄²⁻ concentration data for the 90 and 100 cm depths during the winter were lost due to damage to the incubation chambers.

2.5. Measurement of environmental variables

During each sampling campaign, temperature (°C), pH, and electrical conductivity (EC; mS cm⁻¹) in the top 15 cm soils were measured at each site. Soil temperature and pH were determined *in situ* by using a handheld pH/mV/temperature meter (IQ150, IQ Scientific Instruments, Carlsbad, CA, USA), and soil EC was measured with a EC Meter (2265FS, Spectrum Technologies Inc., Phoenix, AZ, USA). Air temperature (°C) and rainfall were recorded by an automatic meteorological station (LSI-LASTEM, Italy) installed at the Min River Estuary Station of the China Wetland Ecosystem Research Network.

2.6. Data analysis and model formation

Data were log-transformed to approximate normal distributions when selected attributes were skewed. The coefficients of variation (CV) for CH₄ fluxes and environmental variables were calculated by dividing the standard deviation by the mean to determine the magnitude of interannual (among the 5 years) and interseasonal variability (among the 20 seasons observed) (Musavi et al., 2017). Two-way analysis of variance (ANOVA) was used to explore whether seasonality, soil depths or their interaction have fixed effects on soil CH₄ production rates or

porewater CH₄ concentrations, with soil sulfate (SO₄²⁻) concentration being a covariate.

We recognised that the above formed statistical models in this study might not fit the assumptions of ANOVA, rendering the formal inference based on the p-value of ANOVA potentially unreliable. Apart from the ANOVA models, different mixed-effect models were also used to investigate how soil depths were related to soil CH₄ production rates or porewater CH₄ concentrations, because it would be more feasible to model the variance structure of soil depths in the mixed-effect model framework than ANOVA. Since the different models for soil depths were not nested, likelihood ratio tests could not be used, and the Akaike information criterion (AIC) was used instead for model comparison.

To take into account the possible spatial autocorrelations of soil CH₄ production rates or porewater CH₄ concentrations down the soil profile, we also considered soil depth as a random effect variable in the linear mixed-effect model using the lme function in the nlme package of R (Pinheiro et al., 2017). Our results showed that the AIC values of models fitting soil depth as a fixed factor for both soil CH₄ production rates and porewater CH₄ concentrations (158 and 55, respectively) were significantly lower than those fitting soil depth as a random effect variable (211 and 90, respectively). Hence, we only presented results obtained from the former models fitting soil depth as a fixed factor, which performed slightly better than the mixed linear model.

Linear mixed-effect models were also used to test for differences in interseasonal variability of CH₄ fluxes within sites after accounting for the possible effects of air temperature, soil temperature, precipitation, soil pH and water salinity, with sampling year being fitted as a random intercept to account for the repeated measures of other factors, i.e. interseasonal variability ~ air temperature + soil temperature + precipitation + soil pH + salinity, random = ~ 1 | Year. Similarly,

linear mixed-effect models were used to test for the possible predictors of the variations in CH₄ flux, with the sampling site being selected as a random effect variable to account for the repeated measures in spatial CH₄ flux i.e., CH₄ flux ~ air temperature + soil temperature + precipitation + soil pH + salinity, random = ~ 1 | site. In order to test for temporal autocorrelation, we plotted autocorrelation function (ACF) and partial autocorrelation (PACF) plots of the residuals to help interpretation of the CH₄ flux data. Following Bader et al. (2013), we refitted a model including an autocorrelation function with a first-order autoregressive correlation structure (AR1), specified as “correlation = corAR1 (form=~ date | site)”, to account for the repeated measures on 60 different days at the three sites to model the violation of independence of residuals from different sampling days. Significant difference between models with and without AR1 was tested by the anova function in R, and the model with AR1 that showed a significantly lower AIC value was chosen. A variable selection with the fitted global models based on the AIC algorithm and a relative importance method were then used to quantify the contributions of the best predictors (the significant variables of the final model) of the variations in CH₄ flux and their interseasonal variability. For the model selection, we used the stepAIC() function in the R package “MASS”, accompanied by the calc.relimp() function with Lindeman-Merenda-Gold (LMG) relative importance method in the R package “relaimpo” (Musavi et al., 2017). The model with the lowest AIC value was chosen, and the relationship between the dependent variables and chosen predictors was further tested by Type II Wald’s test implemented in the R package “car”.

Besides, the differences in the mean values of environmental variables (precipitation, temperature, soil pH, and soil EC) over the five years were also examined by repeated measures analysis of variance (RMANOVA). For the dataset of each individual year, Pearson correlation

analysis was used to examine the relationships (1) between environmental variables and CH₄ emissions, soil CH₄ production rates, or porewater CH₄ concentrations, and (2) between CH₄ emissions and soil CH₄ production rates or porewater CH₄ concentrations. The interseasonal variability (ISV) of salinity and CH₄ fluxes was determined by dividing the standard deviation of the variables measured at triplicate sampling sites by the average value obtained in each individual season. Temperature sensitivity (Q_{10} value) of CH₄ emissions was calculated following the exponential regression model described by [Tong et al. \(2015\)](#) and [Wang et al. \(2015\)](#). All statistical analyses were performed using R version 3.4.1 ([R Development Core Team 2017](#)) and a P value of < 0.05 was considered statistically significant for multiple comparisons. All data were reported as mean \pm 1 standard error (SE). All statistical graphs were generated using OriginPro 7.5 (OriginLab Corp. USA).

3. Results

3.1. Temporal variations in environmental variables

[Figure 2](#) shows the temporal variations in soil temperature, pH, and EC, all of which showed similar patterns over time for the majority of the study periods. Considerably higher air and soil temperatures and lower EC were observed between May and September than in other months ([Fig. 2](#)). Soil pH showed no clear seasonal patterns but varied slightly among measurement events. Over the five-year period, the mean annual air temperatures were very close to the historical average of 22.2 °C ([Table 1](#)) while the monthly mean air temperatures followed the long-term historical patterns, with July and August usually being the warmest months and January and February the coldest ([Fig. 2](#)). [Fig. S1](#) shows the monthly precipitation amounts over the five study years, which varied significantly both seasonally and inter-annually. Nearly half of

the annual rainfall occurred in summer, with several heavy rainfall events in July and August. Significantly higher annual precipitation was observed in 2013, while lower precipitation occurred in 2007 and 2009.

3.2. Dynamics of soil CH₄ production rates and porewater CH₄ concentrations

The range of average soil CH₄ production rates across all depths among the four seasons was large, spanning three orders of magnitude from 0.04 to 1.67 $\mu\text{g CH}_4 \text{ g}^{-1} \text{ day}^{-1}$. Soil CH₄ production rates varied significantly with season and soil depth ($P < 0.05$) (Table 2 and Fig. 3). The highest and lowest soil CH₄ production rates were observed in the summer and winter, respectively (Table 2 and Fig. 3). Significantly higher CH₄ production rates were observed from the topsoil (5–15 cm) as compared to other soil depths during the spring, summer, and autumn ($P < 0.05$) (Table 2 and Fig. 3), indicating a decreasing trend with depth. There were also significant interactions between seasons and soil depths in affecting soil CH₄ production rates ($P < 0.05$) (Table 2).

Fig. 4 shows the seasonal variations in dissolved CH₄ concentrations down the soil profile. Porewater CH₄ concentrations varied significantly with both depth and season ($P < 0.01$) (Table 2), ranging between 2 and 457 $\mu\text{mol L}^{-1}$ (Fig. 3). We observed a significant, increasing trend of porewater CH₄ concentrations with depths (Table 2 and Fig. 4), and substantially higher CH₄ concentrations at all depths during the summer ($P < 0.01$) (Table 2 and Fig. 4).

3.3. Temporal variations in CH₄ emissions

3.3.1. Seasonal variations in CH₄ emissions

Across all years, the highest CH₄ emissions were observed between April and October (Fig. 5). Fluxes were generally low between November and March, except in 2013 in which the peak

of CH₄ emission occurred in December and January. When averaging the monthly fluxes over five years, a strong seasonal pattern in CH₄ emissions emerged, with generally low values in spring, a maximum in summer, and a minimum in winter (Fig. 6). Meanwhile, we observed considerable variations in both mean CH₄ fluxes (Table 1) and the timing of maximum emissions (Fig. 5) among different years. For example, the maximum CH₄ emissions occurred in May–June in 2013, but in August–October in 2014. Clear peaks of CH₄ emissions were not observed in 2007 and 2009, with only slightly higher fluxes being detected between April and October. Salinity showed clear links to the seasonal variability of CH₄ emissions (Table 3), with a significant negative correlation observed between the two (Table 4). When considering the variability in the CH₄ emission (SD divided by mean or CV), it was positively correlated with the equivalent variability in salinity (Fig. 7); i.e. emissions varied more when also salinity was variable among measurements.

3.3.2. Inter-annual variations in CH₄ emissions

The coefficient of variation of annual mean CH₄ emissions over the five years was 67%, which implied a considerable inter-annual variability. Over the study period, the mean annual CH₄ emissions from the *C. malaccensis* marsh ranged between 0.71 and 5.10 mg CH₄ m⁻² h⁻¹, leading to annual cumulative emissions of 6.2–48.9 g CH₄ m⁻² (Fig. 5 and Table 1). Significantly lower and higher CH₄ effluxes were observed in 2007 and 2013, respectively, as compared to other years (Table 1). According to the AIC-based model selection, variations in CH₄ emissions were best predicted by soil temperature, precipitation and salinity (represented by EC) (Table 3), which independently explained 60.0% (positive effect), 21.7% (positive effect) and 18.2% (negative effect) of the variations, respectively (Fig. 7).

4. Discussion

4.1. Variability of soil CH₄ production rates and porewater CH₄ concentrations

Soil CH₄ production rates from our estuarine marsh demonstrated significant variations down the soil profile (Table 2 and Fig. 3), with the highest rates occurring in the top soil layer (5–15 cm depth) in all seasons except winter, which was in accordance with the results of previous studies (van den Pol-van Dasselaar & Oenema, 1999; Liu et al., 2011; Knoblauch et al., 2015). We found a negative correlation between soil CH₄ production rates and porewater SO₄²⁻ concentrations along the soil profile (Fig. S2). The higher porewater SO₄²⁻ concentrations in the deeper soil layer can help the sulfate-reducing bacteria in outcompeting the methanogens for substrates, thereby inhibiting CH₄ production at depth (van der Gon et al., 2001; Purdy et al., 2003; Vizza et al., 2017). The vertical distribution of CH₄ production rates down the soil profile might also be related to the differences in substrate quantity and quality. Previous studies in wetlands have shown that soil CH₄ production rate increased with the availability of labile carbon fractions (Updegraff et al., 1995; Liu et al., 2011; Inglett et al., 2012). A previous study conducted at our site has shown that the majority of *C. malaccensis* root biomass was distributed in the upper surface layer (Tong et al., 2011), which could provide an abundant supply of labile carbon to support the metabolic activity of methanogens (Ström et al., 2012). On the other hand, we found a significant increase in porewater CH₄ concentrations with depth, which was opposite to the pattern of CH₄ production rates in the soil profile (Table 2 and Fig. 4). The concentration of CH₄ in porewater is influenced by both CH₄ production and loss. In spite of a high CH₄ production rate in the top soils, we hypothesize that the lower porewater CH₄ concentration observed could be related to the tidal actions, which are one of the key physical processes

shaping the biogeochemical processes in coastal wetlands (Tong et al., 2010). The top soil layers were subjected to frequent tidal flushing, which could enhance CH₄ export to the tidal waters and reduce the accumulation of CH₄ in porewater (Lee et al., 2008). In addition, the inflow of tidal water would bring along a large amount of oxygen and SO₄²⁻ to the surface soils, thereby increasing the soil redox potential and promoting methanotrophy in the upper layers (Ding et al., 2003; Sun et al., 2013).

We observed distinct seasonal variations in soil CH₄ production rates with significantly higher values in the summer (Table 2 and Fig. 3), which were in accordance with the findings of previous studies (Bergman et al., 2000; Avery et al., 2003; Tong et al., 2012). Similarly, porewater CH₄ concentrations were found to be significantly higher during the summer season. It is generally acknowledged that CH₄ production rates would vary seasonally as a function of temperature (e.g. Segers, 1998; Inglett et al., 2011). In our study, soil temperature had an exponential relationship with soil CH₄ production rates (Fig. S3), and positive correlation with porewater CH₄ concentrations ($r = 0.662$, $p < 0.01$, $n = 24$), pointing to the positive impacts of temperature on microbial-mediated methanogenesis. Moreover, the amount of plant biomass in this wetland was found to be much higher in summer than in winter (Tong et al., 2011). The enhanced plant productivity and subsequently supply of labile carbon substrates through root exudation in the summer period would likely play a role in stimulating methanogenic activities (Whiting & Chanton, 1993; Bergman et al., 2000; Walter et al., 2001) and hence increasing the concentrations of CH₄ in soil porewater (Xiang et al., 2015). In addition, the increased freshwater discharge from the estuary in summer time provided a dilution effect that significantly reduced the salinity of tidal water, which would in part facilitate methanogenesis through reduced

competition with sulfate-reducing bacteria (Sinke et al., 1992). We observed a significant and negative correlation between salinity and porewater CH₄ concentration ($r = -0.653$, $p < 0.01$, $n = 24$) that supported this hypothesis.

4.2. Temporal variations of CH₄ emissions

4.2.1. Seasonal variability

In this study, CH₄ emissions from the subtropical estuarine marsh varied considerably among different seasons. The seasonal mean CH₄ emissions over the five-year period were correlated significantly with both soil CH₄ production rates (0–20 cm depth) (Fig. S4) and porewater CH₄ concentrations (Fig. S5). As such, the seasonal pattern of CH₄ emission (Fig. 6) was highly similar to that of soil CH₄ production rates (Fig. 3) and porewater CH₄ concentrations (Fig. 4). This strong relationship was expected since a high CH₄ production rate in soils would increase the supply of CH₄ to soil porewater, and subsequently enhance net CH₄ emissions to the atmosphere owing to the steeper concentration gradient.

The seasonal variability of CH₄ emissions could be governed by the interactions of a number of environmental variables. Our results showed that salinity was clearly and negatively correlated with the variations of CH₄ flux among different seasons (Table 3 and Table 4). Numerous studies have reported a significant reduction in CH₄ emissions from coastal wetlands with salinity (Bartlett et al., 1987; Magenheimer et al., 1996; Poffenbarger et al., 2011; Tong et al., 2012; Sun et al., 2013; Vizza et al., 2017). The significantly lower soil salinity (represented by EC) observed between May and September in our site could significantly enhance methanogenic activities owing to reduced presence of alternate electron acceptors (Welti et al., 2017). Salinity could also affect CH₄ production through its effects on extracellular enzyme activities and carbon

mineralization rates (Chambers et al., 2013; Neubauer et al., 2013). Meanwhile, salinity might also affect methanotrophic activities directly or indirectly, which in turn alter the rate of CH₄ emissions from wetlands. Only few studies have thus far directly examined the mechanistic processes, i.e. CH₄ production and oxidation, involved in the suppression of net CH₄ flux by salinity (Vizza et al., 2017). Further studies should be carried out to explore the exact impacts of salinity on various biogeochemical processes in soils in affecting CH₄ dynamics.

Temperature was another important predictor of the changes in CH₄ emissions from our *C. malaccensis* marsh, as shown by the strong relationships observed between air/soil temperature and CH₄ flux in individual years (Table 4). An increase in temperature could enhance CH₄ emissions by increasing methanogenic activities, stimulating root exudations (Song et al., 2009; Yvon-Durocher et al., 2014; Olsson et al., 2015), as well as facilitating plant-mediated CH₄ transport (Hosono and Nouchi, 1997). Meanwhile, we found that the temperature sensitivity of CH₄ flux varied considerably among different years over the study period, with $Q_{\text{air}10}$ and $Q_{\text{soil}10}$ values ranging from 2.46 to 5.30, and from 3.66 to 7.92, respectively (Fig. S6). Our results suggest that the estimation of long-term (multi-year) CH₄ emissions based on simple extrapolations of the relationships between temperature and CH₄ flux derived from short-term (< 1 year) measurements might not be reliable and introduce significant biases. Apart from temperature, the hydrologic conditions of the site could also affect CH₄ emissions by controlling the depths of the oxic and anoxic layers as well as soil redox potential (Dinsmore et al., 2009). The disproportionately high amount of precipitation received during the summer (Fig. S1) could favour the formation of a wetter and more anaerobic environment in the soils for methanogenesis (Lai et al., 2014). Furthermore, the total amount of plant biomass (aboveground + belowground)

at our marsh site was found to vary significantly among seasons in the following order: summer > autumn > spring > winter (Tong et al., 2011), which could exert influences on the variability of plant-mediated CH₄ emissions via primary production and substrate supply.

Based on our five-year data set, we observed that peak CH₄ emissions generally occurred during the summer period when temperature was high and conductivity was low, which favored methanogenesis. Yet, the exact timing of peak CH₄ emissions varied from one year to another that could be partly related to the inter-annual variations in the timing of maximum monthly precipitation, which governed the extent of anaerobic conditions in soils. For instance, the timing of peak CH₄ emission coincided with that of maximum monthly precipitation in 2008 and 2014, which happened to be in the months of July and August, respectively. Yet, in 2013, the extremely high precipitation amount in July implied a lack of abundant sunlight during this period, which could hinder photosynthesis by marsh plants and the supply of labile carbon from photosynthates to soils for methanogenesis. Our results point to a need of carrying out more in-depth studies in future to disentangle the specific influences of various environmental factors on the seasonal variability of CH₄ emission in coastal marshes.

4.2.2. Inter-annual variability

In the present study, CH₄ emissions from the brackish *Cyperus malaccensis* marsh showed substantial inter-annual variability (Fig. 5 and Table 3). Previous studies have shown that the inter-annual variability of CH₄ emissions was governed by water table position (Moore et al., 2011), peat temperature (Shannon and White, 1994; Lai et al., 2014), and precipitation (Song et al., 2009). According to the AIC-based model selection, we found that soil temperature and salinity were the primary determinants of the inter-annual variability of CH₄ flux at our site

(Table 3 and Fig. 7a). The effects of soil temperature and salinity could be related to the production of substrate precursors and methanogenic activity as described previously (Whalen et al., 2005; Dinsmore et al., 2009; Lai et al., 2014; Yvon-Durocher et al., 2014). In addition, we found strong correlations between CH₄ flux and precipitation amount over the study period (Table 4). Among the five study years, the lowest annual mean precipitation was recorded in 2007, which was significantly lower than that in 2008, 2013 and 2014 (1362 vs. 1485-1890 mm, $p < 0.05$, Fig. S1). It was likely that the much lower CH₄ emission observed in 2007 was at least partly related to the significantly lower amount of precipitation received in this particularly dry year, which provided a more aerobic and less favorable environment for methanogenesis to take place. Other factors, such as primary productivity and water table depth, might also contribute to the inter-annual variations in CH₄ flux and deserve further investigations.

It is noteworthy that significantly higher CH₄ emissions were observed in 2013 and 2014 as compared to other years ($p < 0.05$; Table 1), which could be related to the increased discharge of nutrient-enriched effluents from the aquaculture ponds in the surrounding region. Starting from 2011, the conversion of natural tidal marshes into aquaculture ponds has become increasingly dominant in the Shanyutan wetland. A previous study estimated that about 29% of total nitrogen and 16% of total phosphorus added to the ponds in the form of feeds and fertilizers were actually assimilated by fish and shrimps during a production cycle (Avnimelech and Ritvo, 2003). The majority of added nitrogen and phosphorus would then eventually be discharged as effluents that are rich in particulate matters (e.g. uneaten feeds, faeces, phytoplankton) and dissolved nutrients (Jackson et al., 2004; Molnar et al., 2013) into the adjacent waterbodies, further stimulating microbially-mediated CH₄ emissions. Studies based on both laboratory incubations and field

measurements have demonstrated the positive effects of exogenous nutrient loading on CH₄ production and emissions (Liu and Greaver, 2009; Chen et al., 2010). Hu et al. (2017) also reported that the addition of nitrogen strongly stimulated CH₄ emissions from the *C. malaccensis* marsh in our study area.

4.3. Implications and further outlook

It is generally considered that coastal wetlands have a lower CH₄ emission rate than other natural wetlands owing to the inhibitory effect of high SO₄²⁻ concentrations (Bartlett et al., 1987; Ding et al., 2004b; Saarnio et al., 2009; Poffenbarger et al., 2011). However, we found that the mean annual CH₄ emission from our subtropical brackish marsh in the Min River Estuary was 23.8±18.1 g CH₄ m⁻² yr⁻¹, which was 1.8 times higher than the average of 13.3 g CH₄ m⁻² yr⁻¹ across China's natural wetlands (Wei and Wang, 2017), and also substantially higher than the annual CH₄ flux reported in the Atlantic blanket bogs (5.5–6.2 g CH₄ m⁻² yr⁻¹) (Laine et al., 2007; Koehler et al., 2011), Finnish bogs (5.2–6.8 g CH₄ m⁻² yr⁻¹) (Alm et al., 1999), as well as Swedish and Minnesota fens (12.0–19.5 g CH₄ m⁻² yr⁻¹) (Shurpali et al., 1993; Rinne et al., 2007; Nilsson et al., 2008). The range of CH₄ emissions from our brackish marsh (6.19–48.86 g CH₄ m⁻² yr⁻¹) was also comparable to that from the coastal wetlands in South India (17.3–118.4 g CH₄ m⁻² yr⁻¹) (Purvaja and Ramesh, 2001), freshwater marshes in Europe (10–90 g CH₄ m⁻² yr⁻¹) (Saarnio et al., 2009), and freshwater marshes in China (4.9–94.1 g CH₄ m⁻² yr⁻¹) (Ding et al., 2004b). Our results suggest that subtropical estuarine brackish marshes could be important sources of atmospheric CH₄ and thus should not be overlooked in greenhouse gas accounting for their contributions to global climate change.

Numerous efforts have been made to estimate regional CH₄ emissions from coastal wetlands

by extrapolating short-term (< 2 year) field measurements to longer periods in a given area (e.g. [Purvaja and Ramesh, 2001](#); [Ding et al., 2004b](#); [Saarnio et al., 2009](#); [Ortiz-Llorente and Alvarez-Cobelas, 2012](#)). However, our results show that CH₄ emissions from the estuarine marshes had strong inter-annual variability, which suggest a high uncertainty of regional estimates of CH₄ emissions relying only on short-term measurements. It is of paramount importance to take into account long-term observations in order to reduce the uncertainty of CH₄ flux estimations and improve our understanding of the impacts of wetlands on the atmospheric CH₄ balance. Moreover, most of the existing process-based models used for estimating CH₄ flux from coastal wetlands fail to consider the influence of salinity ([Li et al., 2016](#)). In this study, we observed a significant and strong negative relationship between salinity and CH₄ emissions ([Table 3 and Table 4](#)), indicating that the interactions between temperature, salinity, and other biotic/abiotic factors should be addressed comprehensively to improve the current CH₄ flux models.

It should be noted that the findings of this study were limited by several uncertainties, which could be associated with the following aspects: (1) single time-point measurements do not fully capture the episodic and high magnitude events of CH₄ release; (2) a limited number of flux measurement sites results in a lack of adequate spatial representation; (3) chamber measurement problems, such as changes in temperature; and (4) the lack of CH₄ emission observations during flood tide. Future research programs will thus need to increase the frequency of sampling *in situ* for longer periods and at different spatial scales as well as include innovative techniques (e.g. eddy covariance tower) in order to measure CH₄ releases from coastal marshes.

5. Conclusions

Our long-term measurements of CH₄ emissions from the subtropical estuarine brackish *C. malaccensis* marsh ecosystem in the Min River Estuary, southeast China over five years revealed strong inter-annual and seasonal variabilities of CH₄ fluxes. The temporal variations in CH₄ emissions from our marsh were mainly related to the variations in soil temperature and salinity, while the potential roles of primary productivity and precipitation should not be overlooked. Our results suggest that long-term, high-frequency observation of CH₄ emissions is essential for making reliable flux estimates from the coastal marshes. In addition, the significant relationships among soil CH₄ production rates, porewater CH₄ concentrations, and net CH₄ emissions observed in this study highlight the great potential in successfully simulating these processes using biogeochemical models once the influences of key environmental factors are properly quantified.

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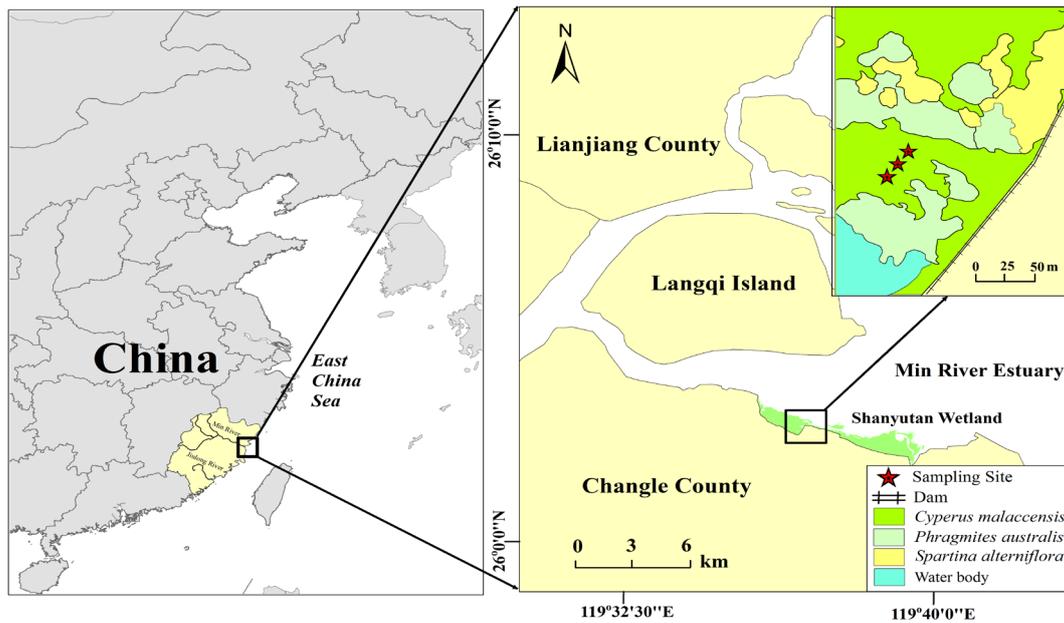
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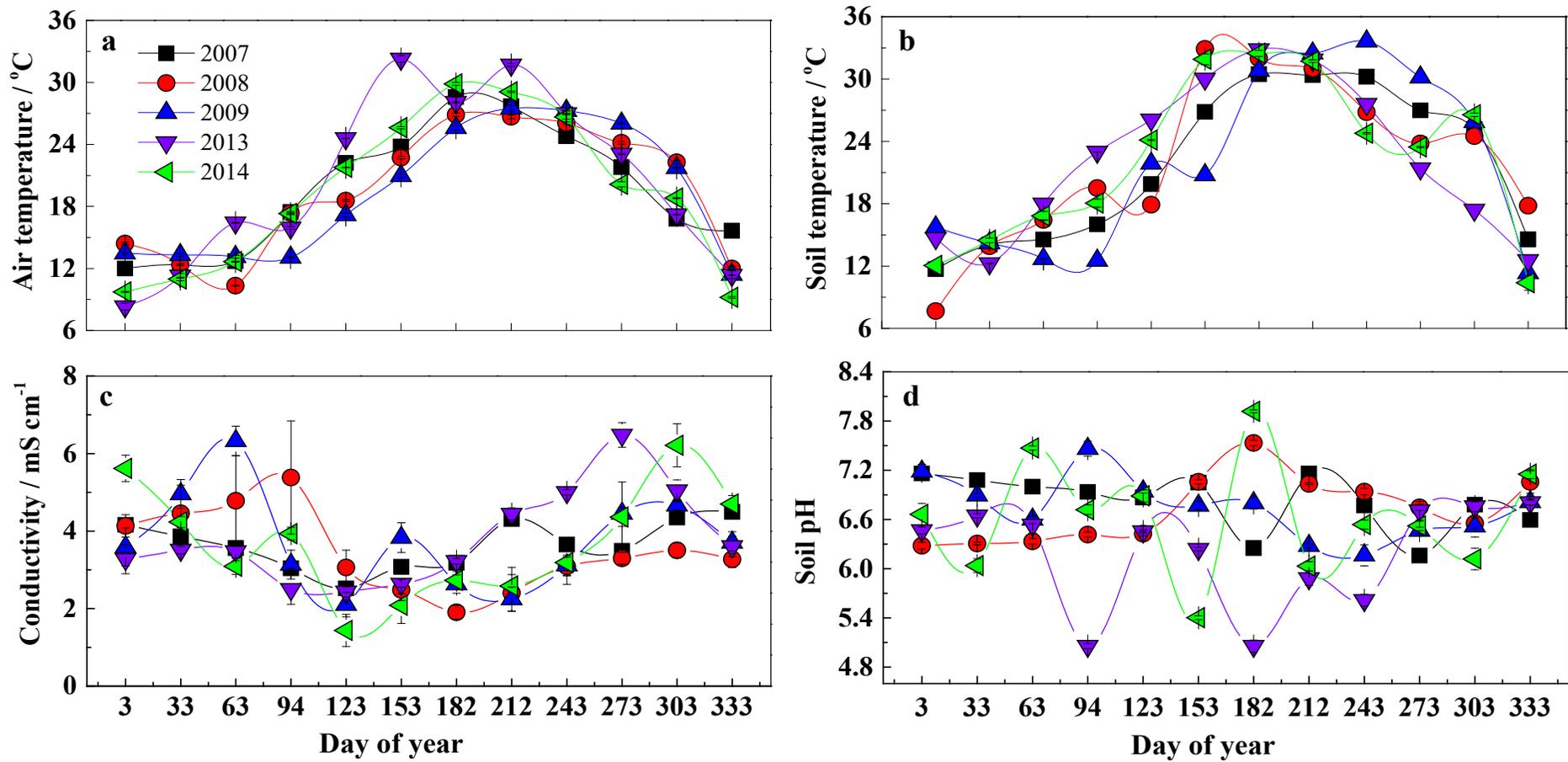
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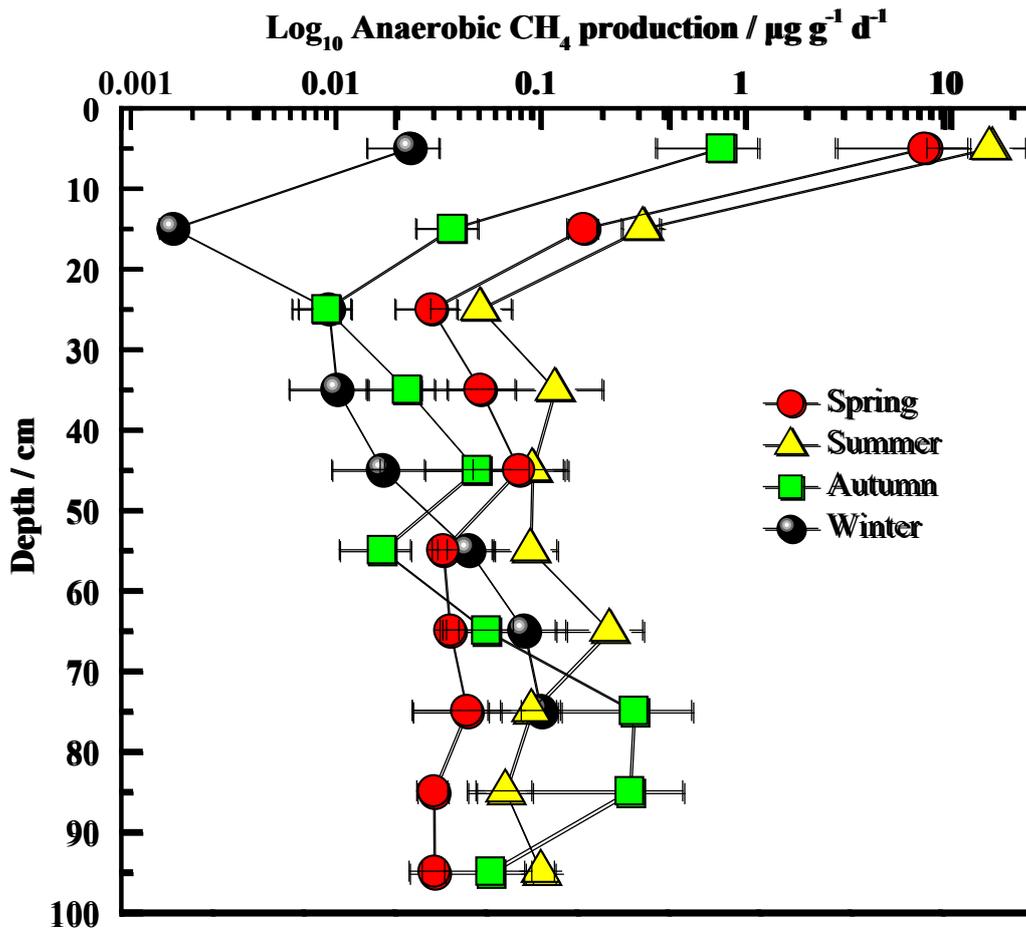
2 **Fig. 1.** Locations of the sampling sites in the subtropical brackish *C. malaccensis*
 3 marsh ecosystem in the Min River estuary.



4

5 **Fig. 2.** Temporal variations in (a) air temperature, (b) soil temperature, (c) soil electrical conductivity, and (d) soil pH at the top 25 cm depth in

6 the brackish *C. malaccensis* marsh over the five-year study period. Values are means \pm 1 S.E. ($n = 3$).

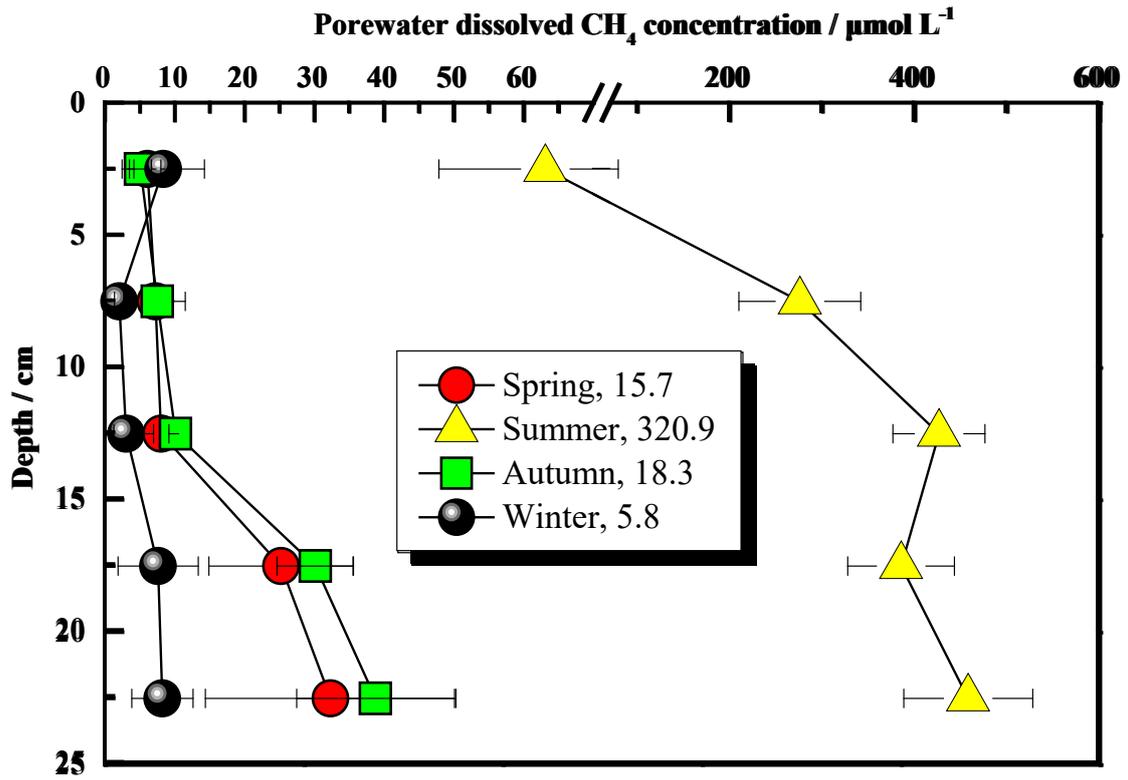


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8 **Fig. 3.** Vertical profiles of CH₄ production rates in the top 100 cm soils of the brackish

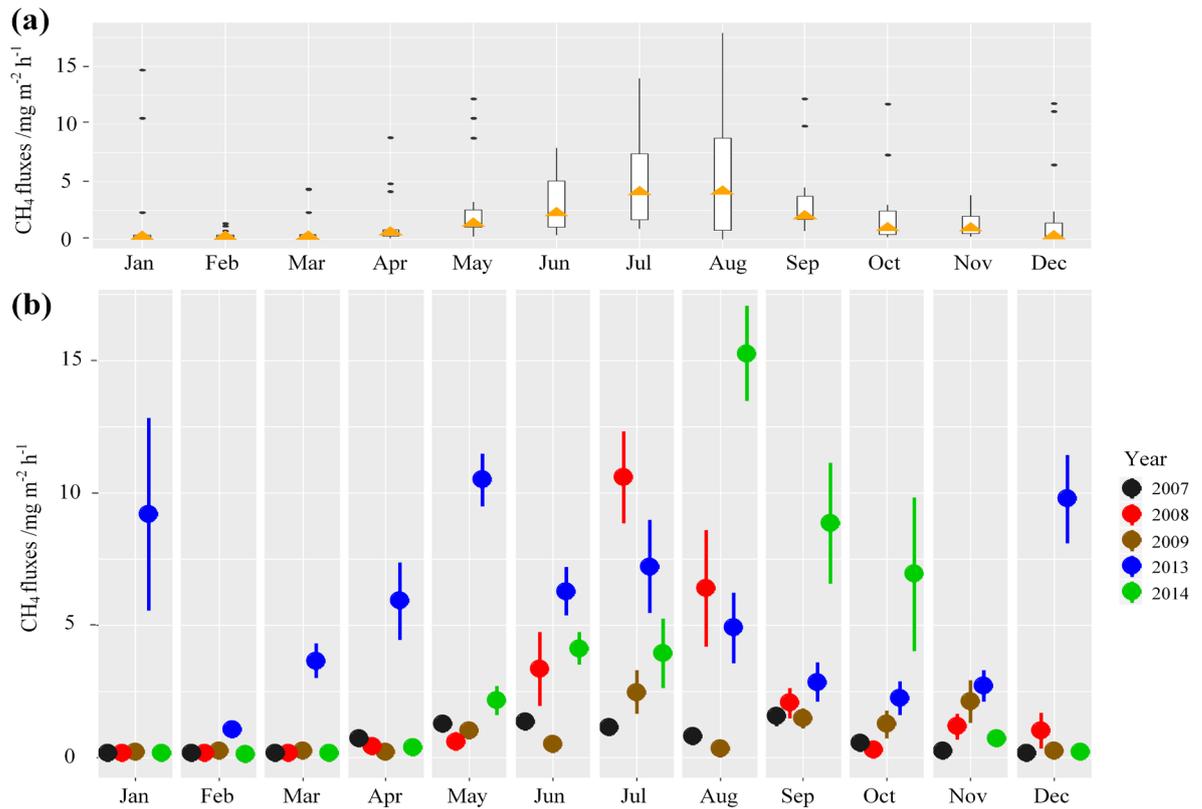
9 *C. malaccensis* marsh among the four seasons in 2012. Values are means ± 1 S.E. (*n* =

10 3).



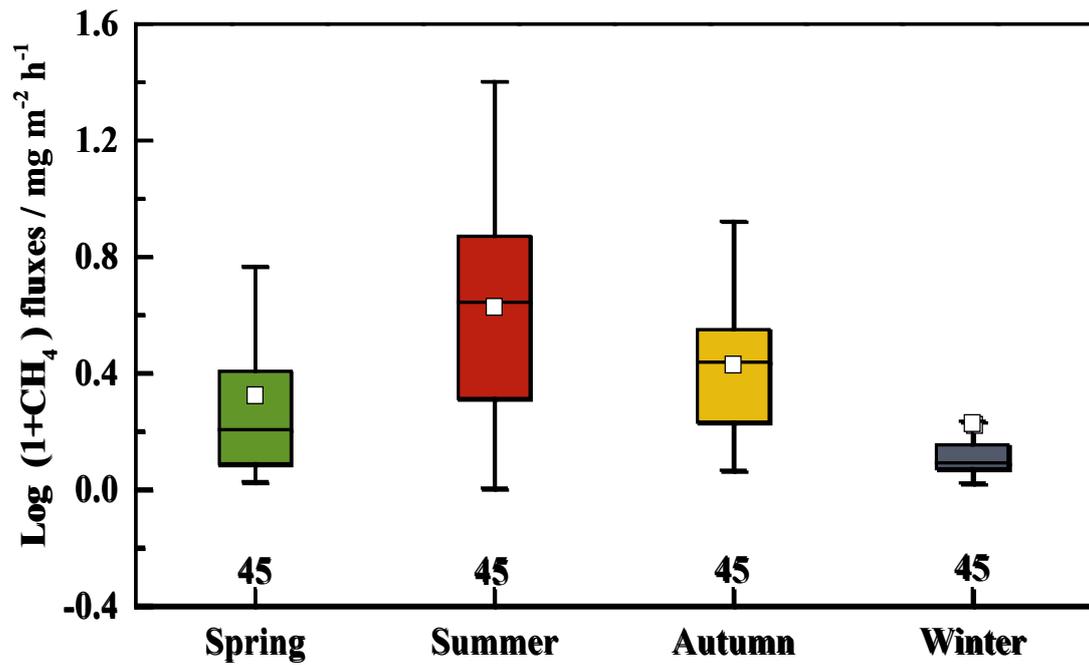
11

12 **Fig. 4.** Seasonal variations in porewater CH₄ concentrations in the brackish *C.*
 13 *malaccensis* marsh in 2012-2013. The numbers next to the symbols in the figure
 14 legend represent the mean porewater CH₄ concentrations in the top 25 cm soils.
 15 Values are means ± 1 S.E. (*n* = 12).



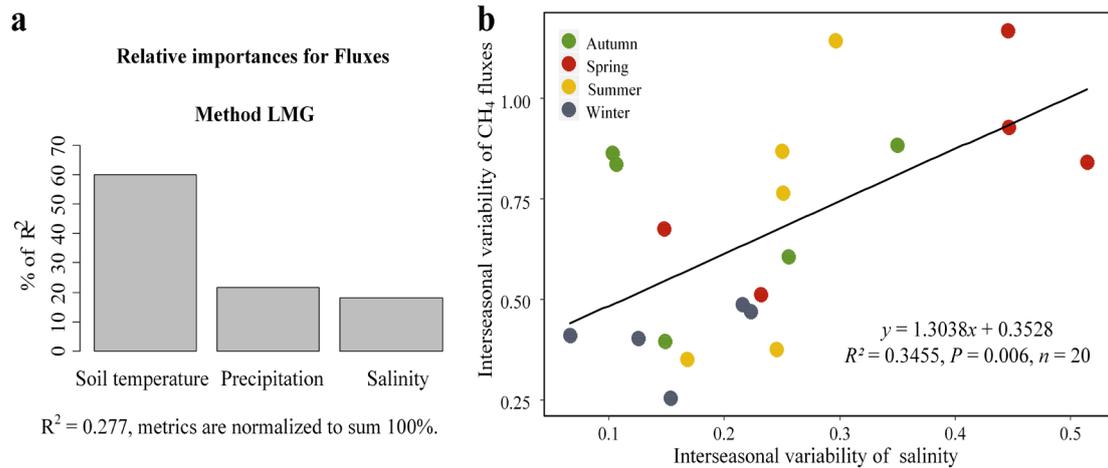
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17 **Fig. 5.** (a) Monthly variations in average CH₄ emissions in the brackish *C.*
 18 *malaccensis* marsh over the five-year period. Box plots show the median (triangle),
 19 25–75th percentile (limits), minimum and maximum values without outliers
 20 (whiskers), and outliers (black dots). (b) Monthly variations in CH₄ emissions in the
 21 brackish marsh in individual years. The dots and bars represent the means and
 22 standard errors, respectively.



23

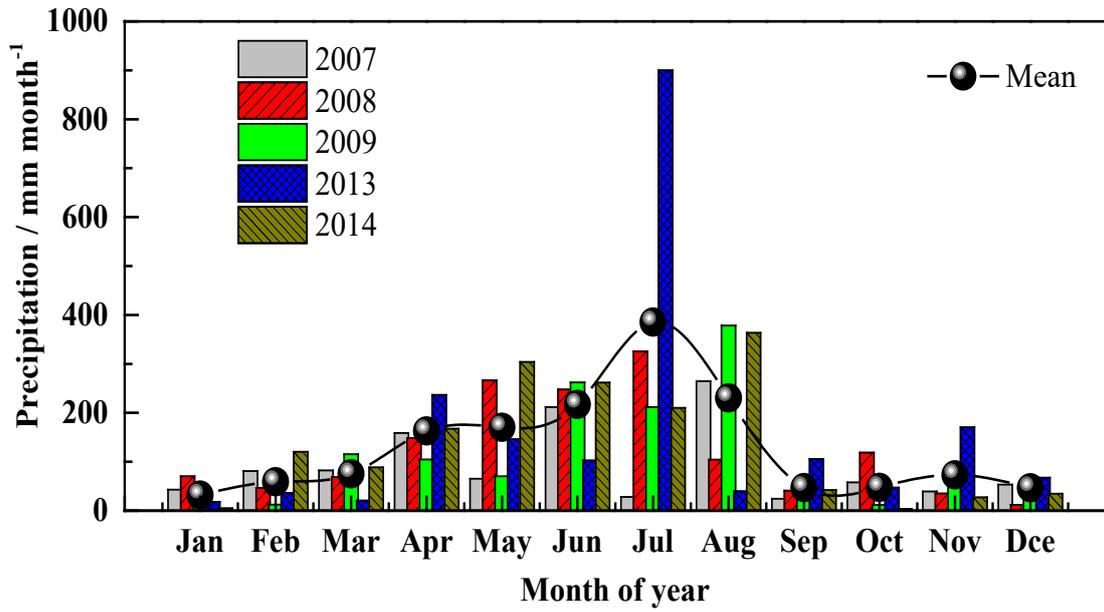
24 **Fig. 6.** Boxplots of seasonal variations in CH₄ emissions in the brackish *C.*
 25 *malaccensis* marsh over the study period. The boxes show the quartiles and median,
 26 while the squares and whiskers represent the means and values within 1.5 times of the
 27 interquartile range, respectively. The numbers above the x-axis labels represent the
 28 number of measurements in each season.



29

30 **Fig. 7.** (a) Relative importance of the three significant factors (soil temperature,
 31 precipitation and salinity) governing CH_4 fluxes. The R^2 value of 0.277 represented
 32 the proportion of variance explained by the fitted model, with the metrics being forced
 33 to sum to 100% using the R package of relaimpo and the command “rela=TRUE”; (b)
 34 The relationship between interseasonal variability of salinity and that of CH_4 fluxes.
 35 The inter-seasonal variability of salinity and CH_4 fluxes was computed by dividing
 36 the standard deviation by the mean.

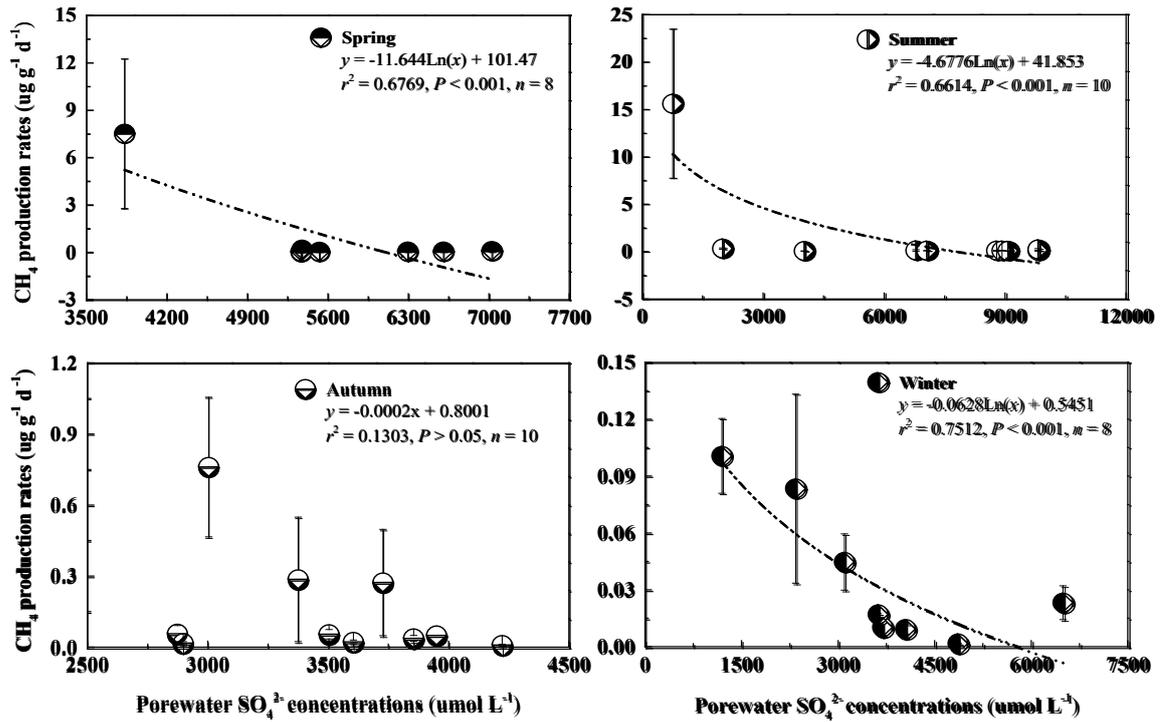
37 **Supporting Information**



38

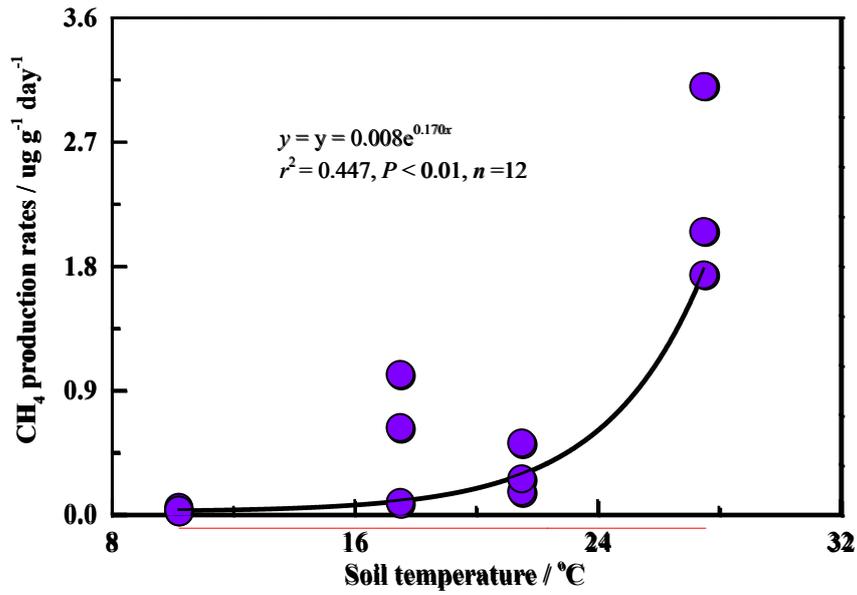
39 **Fig. S1.** Monthly precipitation in 2007–2009 and 2013–2014 for the Min River Estuary Station

40 (mean represents the average for 2007–2009 and 2013–2014).



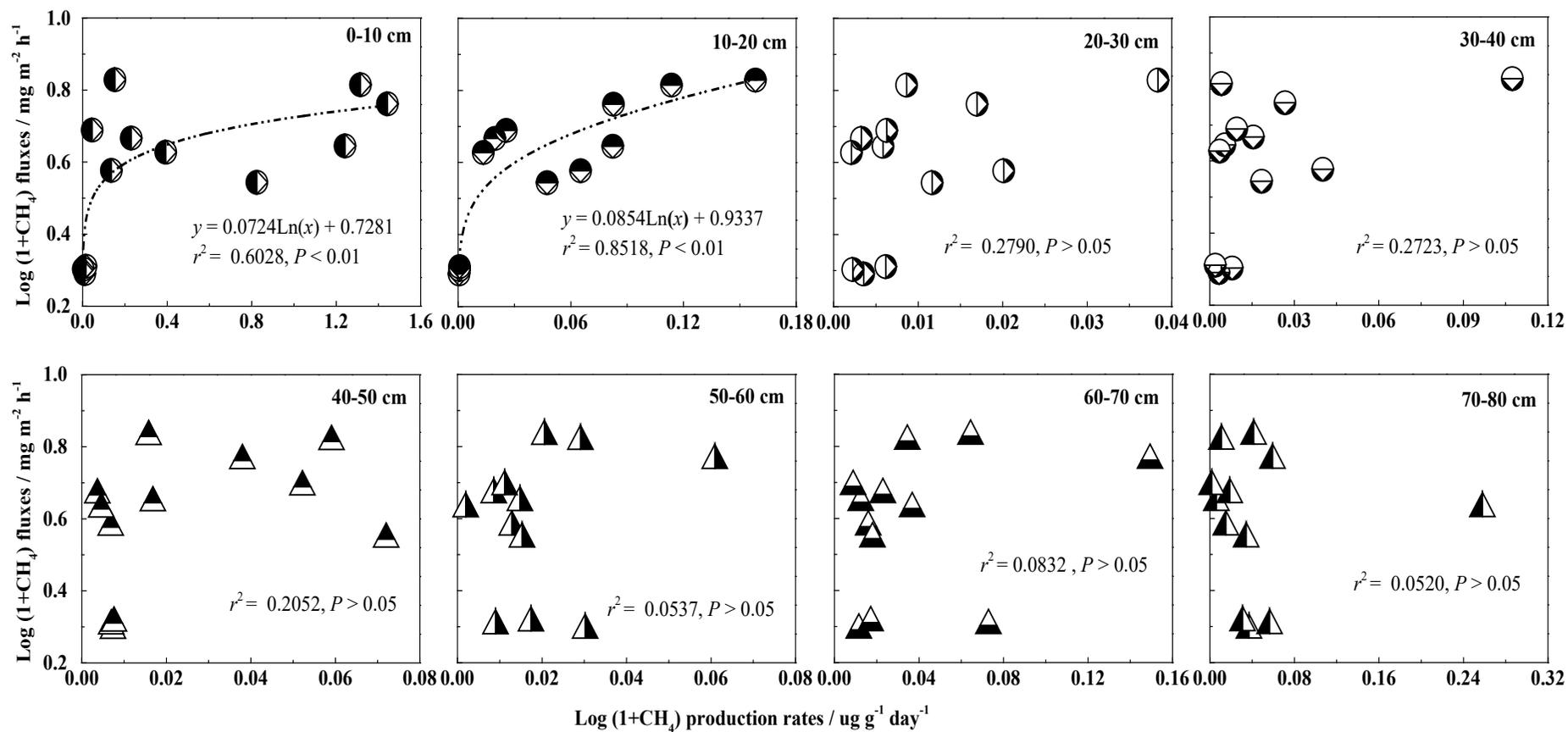
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42 **Fig. S2.** Regression and significance analysis between profiles of CH₄ production rate (y) and
 43 porewater SO₄²⁻ concentration (x) in the brackish *C. malaccensis* marsh for 2012. The soil
 44 porewater SO₄²⁻ concentrations during summer and autumn were for the depth of 0-100 cm ($n =$
 45 10). The soil porewater SO₄²⁻ concentrations during spring and winter were for the depth of 0-80
 46 cm ($n = 8$). CH₄ production rates and porewater SO₄²⁻ concentrations in each soil depth were the
 47 averaged values of three sampling sites.



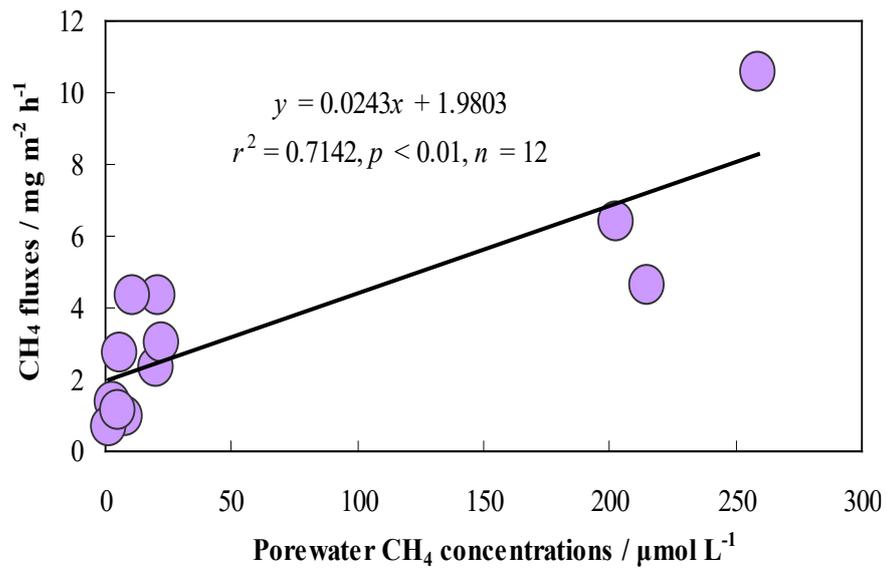
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49 **Fig. S3.** Relationship between the average seasonal CH₄ production rates at five depths and soil
 50 temperature in the brackish *C. malaccensis* marsh for 2012.



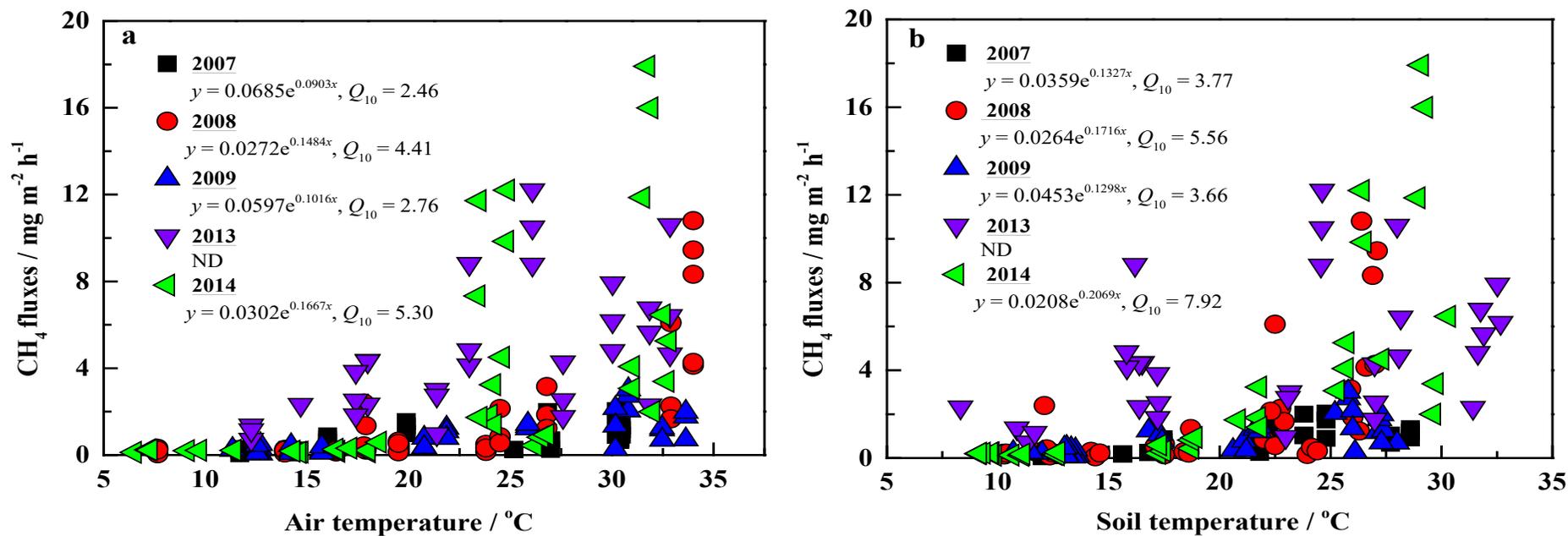
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52 **Fig. S4.** Relationship between seasonal mean CH₄ emissions and soil CH₄ production rate at different depths in the brackish *C. malaccensis* marsh.



53

54 **Fig. S5.** Relationship between seasonal mean CH₄ emissions and mean porewater CH₄
 55 concentration in the brackish *C. malaccensis* marsh for 2013. (12 samples = 1 average value at
 56 five depths × 3 sites × 4 seasons).



57

58 **Fig. S6.** Relationships between CH₄ emissions and air temperature (°C) or soil temperature at the surface soil (0–25 cm) in the brackish *C. malaccensis* marsh from
 59 2007 to 2008 and 2013 to 2014, as described by the exponential function ($P < 0.05$). $n = 36$ for air temperature, soil temperature and CH₄ emissions of each year. ND
 60 means “no significant relationship”.

61

1 **Table 1**

2 Mean, minimum, and maximum values for the selected environmental variables and
 3 CH₄ emissions in the brackish *C. malaccensis* marsh from 2007 to 2008 and 2013 to
 4 2014.

	2007	2008	2009	2013	2014
Air temperature / °C					
Minimum ^a	11.70±0.00	7.66±0.06	12.53±0.00	12.26±0.00	10.40±0.72
Mean ^b	21.74±2.11	22.02±2.26	21.84±2.45	22.30±2.14	22.24±2.25
Maximum ^a	30.47 ±0.00	32.90±0.00	33.63±0.03	32.9±0.00	32.5±0.00
Soil temperature / °C					
Minimum ^a	12.00±0.00	10.33±0.03	11.40±0.38	8.31±0.33	9.20±0.08
Mean ^b	19.65±1.72	19.48±1.76	19.22±1.82	20.60±2.39	19.32±2.17
Maximum ^a	28.60±0.00	26.87±0.15	27.50±0.26	32.27±0.32	29.86±0.15
Conductivity / mS cm⁻¹					
Minimum ^a	2.52±0.00	1.91±0.18	2.10±0.32	2.43±0.13	1.44±0.42
Mean ^b	3.64±0.18	3.48±0.30	3.73±0.35	3.80±0.35	3.68±0.41
Maximum ^a	4.50±0.00	5.38±1.46	6.33±0.38	6.48±0.32	6.22±0.56
pH					
Minimum ^a	6.25±0.00	6.28±0.04	6.16±0.13	5.06±0.03	5.40±0.02
Mean ^b	6.82±0.10	6.72±0.12	6.74±0.11	6.19±0.18	6.62±0.20
Maximum ^a	7.16±0.00	7.53±0.03	7.46±0.09	6.76±0.06	7.92±0.06
CH₄ flux / mg m⁻² h⁻¹					
Minimum ^a	0.19±0.07	0.19±0.07	0.22±0.08	1.07±0.19	0.20±0.04
Mean ^b	0.71±0.15	2.50±1.17	0.87±0.23	5.10±0.60	3.86±1.44
Maximum ^a	1.56±0.34	13.99±5.11	2.47±0.82	10.49±0.98	15.92±2.80
Cumulative emissions / g CH₄ m⁻²					
	6.19	22.14	7.68	48.86	34.13

5 ^a Values are means (±S.E.) of samples (*n* = 3) measured from the brackish *C. malaccensis* marsh
 6 over all sampling sites. ^b Values are means (±S.E.) of samples (*n* = 36) measured from the brackish
 7 *C. malaccensis* marsh over all sampling sites and sampling periods.

8 **Table 2**

9 Summary of two-way analysis of variance ANOVA (season and depth as factors and the sulfate (SO₄²⁻) concentrations as covariates) models
 10 fitted the effect of season, soil depth and their interaction on the CH₄ production rates and porewater CH₄ concentrations in the *C. malaccensis*
 11 marsh.

	CH ₄ production rate					Porewater CH ₄ concentration				
	<i>df</i>	Sum of squares	Mean square	<i>F</i> values	<i>P</i> values	<i>df</i>	Sum of squares	Mean square	<i>F</i> values	<i>P</i> values
Season	3	9.11	3.0366	15.918	<0.001	3	29.366	9.789	90.2	<0.001
Soil depth	9	11.963	1.3292	6.968	<0.001	4	3.533	0.883	8.139	<0.001
SO ₄ ²⁻ concentration	1	2.412	2.4119	12.643	<0.001	1	0.048	0.048	0.439	0.512
Season × soil depth	23	10.232	0.4448	2.332	0.003	12	1.777	0.148	1.365	0.224
Residuals	71	13.544	0.1908			39	4.232	0.109		

12 Note: both the data of the CH₄ production rates and porewater CH₄ concentrations were log-transformed to meet the homogeneity of variances.

13 **Table 3**

14 **(a)** Summary of linear mixed model fitted for CH₄ fluxes accounting for random site effect and temporal autocorrelations, and **(b)** summary of
 15 linear mixed model fitted for interseasonal variability of CH₄ fluxes accounting for repeated measurement among different years. Models are
 16 ranked in order of the lowest Akaike information criterion corrected for low samples sizes (AIC) along with delta AIC. The predictors of the best
 17 model with lowest AIC were tested by Type II Wald test and the significant positive (↑) or negative effects (↓) of chosen continuous predictors
 18 are indicated.

	AIC	delta AIC	Step	df	Deviance	Residual df	Residual deviance
(a)							
CH₄ fluxes							
Soil temperature+ Precipitation +Salinity	907.28	0.00	- Air temperature	1	1.30	173	860.84
Air temperature+Soil temperature+Precipitation+Salinity	912.62	5.34	- pH	1	0.36	172	859.54
Air temperature+Soil temperature+Precipitation+pH +Salinity	918.89	11.61	- Year	4	13.97	171	859.18
Year+Air temperature+Soil temperature+Precipitation+pH+Salinity	931.45	24.17	- Season	3	1.68	167	845.20
Year+Season+Air temperature+ Soil temperature+Precipitation+pH+Salinity	949.68	42.4				164	843.52
<i>Predictors from best model tested</i>						F values	P values
Soil temperature (↑)				1		17.36	<0.001
Precipitation(↑)				1		8.58	<0.001
Salinity (↓)				1		5.92	0.0149
(b)							
Interseasonal variability	AIC	delta AIC	Step	df	Deviance	Residual df	Residual deviance
Salinity _{cv}	21.73	0.00	- Air temperature _{cv}	1	2.36	16	-4.80
Air temperature _{cv} +Salinity _{cv}	26.01	4.28	- pH _{cv}	1	1.72	15	-7.16

Air temperature _{cv} +pH _{cv} +Salinity _{cv}	30.93	9.20	- Precipitation _{cv}	1	0.82	14	-8.88
Air temperature _{cv} +Precipitation _{cv} +pH _{cv} +Salinity _{cv}	36.74	15.00	- Soil temperature _{cv}	1	0.47	13	-9.70
Air temperature _{cv} +	42.91	21.18				12	-10.17
Soil temperature _{cv} +Precipitation _{cv} +pH _{cv} +Salinity _{cv}							
<i>Predictors from best model tested</i>						F values	P values
Salinity _{cv} (↑)				1		10.56	0.001

19 **Table 4**

20 Pearson correlation analysis between CH₄ emissions and environmental variables in
 21 the brackish *C. malaccensis* marsh^a. NS means “no significant relationship”. Bold
 22 numbers denote correlation coefficients for significant relationships.

Environmental variables	CH ₄ fluxes (mg m ⁻² h ⁻¹)					
	2007	2008	2009	2013	2014	All years
Precipitation (mm month ⁻¹)	0.339*	0.517**	NS	NS	0.403*	0.286**
Air temperature (°C)	0.613**	0.609**	0.750**	NS	0.581**	0.373**
Soil temperature (°C)	0.738**	0.510**	0.711**	NS	0.680**	0.349**
Soil pH	NS	0.679**	-0.427**	NS	NS	NS
Soil conductivity (mS cm ⁻¹)	-0.572**	-0.409*	-0.384*	-0.456**	-0.334*	-0.305**

23 ^a *n* = 36 for environmental variables and CH₄ emissions of each year from the *Cyperus*
 24 *malaccensis* marsh. The symbols * and ** indicate significant correlations at the 0.05 and 0.01
 25 levels, respectively.

26

Supporting Information

Title: Methane dynamics in an estuarine brackish *Cyperus malaccensis* marsh: Production and porewater concentration in soils, and net emissions to the atmosphere over five years

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Supporting Information Summary

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Materials and Methods

Methane gas sampling and flux estimation

The CH₄ flux (F , mol m⁻² h⁻¹) was calculated according to the following equation (1):

$$F = \frac{dn}{dt} \times \frac{1}{A} \quad (1)$$

where dn/dt is the slope of amount of substance for CH₄ over sampling period (mol h⁻¹); A is the chamber area (m²). The amount of CH₄ in the chamber at different times was calculated from the following equation (2):

$$n = ppm \times 10^{-6} \times \frac{P_{tot} \times V}{R \times T} \quad (2)$$

where *ppm* is parts per million (usually comes from GC measurement); P_{tot} is total air pressure (usually around 1 atm = 1013.15 hPa); V is the chamber volume (L); R is common gas constant (0.082056 L atm K⁻¹ mol⁻¹); T is the absolute temperature during sampling (K). In present study, the unit of CH₄ flux was showed as mg m⁻² h⁻¹ ($= F \times M \times 10^3$), M is molar mass of CH₄ (g mol⁻¹).

Collection and analysis of SO₄²⁻ concentrations in porewater

To determine porewater SO₄²⁻ concentrations across different soil depths, triplicate sediment cores down to 100 cm depth were collected in January (winter), March (spring), July (summer) and October (autumn) of 2012. The sediment cores were collected using a steel sediment samplers ($\Phi = 5$ cm) and split into ten depths at 10 cm intervals. Soil samples were immediately placed into valve bag and sealed. These sediment samples were subsequently kept on ice in coolers, and transported to the laboratory within 6 h. Upon return to the laboratory, porewater from each depth interval was extracted from the sediment by centrifugation (5000 rpm, 10 min, Cence® L550) and then filtered (0.45 μm acetate fibre membranes). Finally, the SO₄²⁻ concentration was determined using the barium chromate colorimetric method. The incubation chambers equipped with the soil depth of 90 and 100 cm fall to the ground and damaged during the winter experimental period. Therefore, the soil SO₄²⁻ concentration at 90 and 100 cm during the winter is not present in this study.

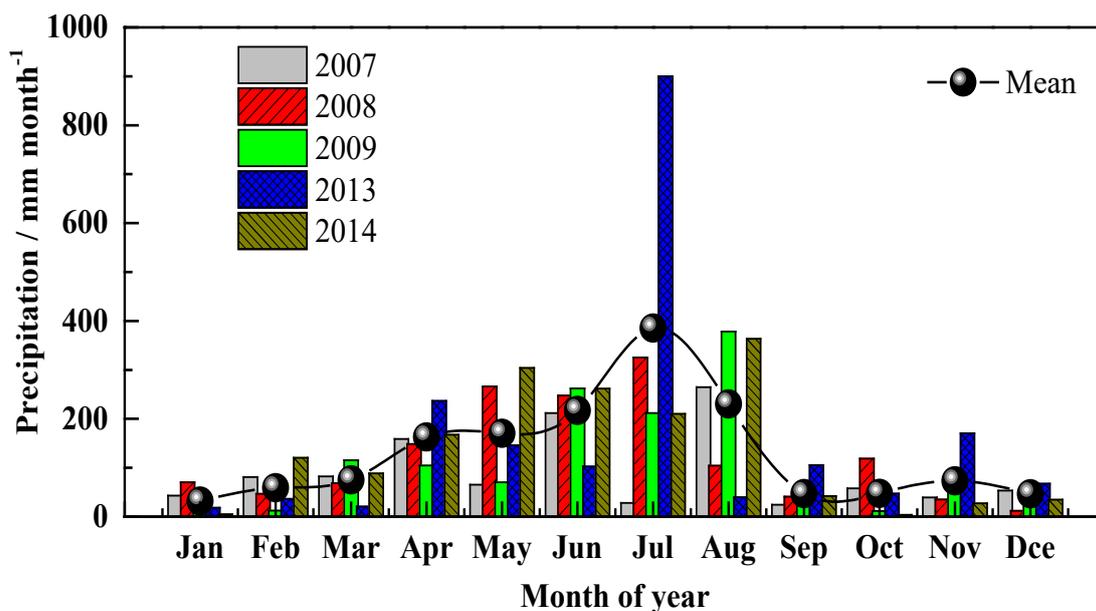


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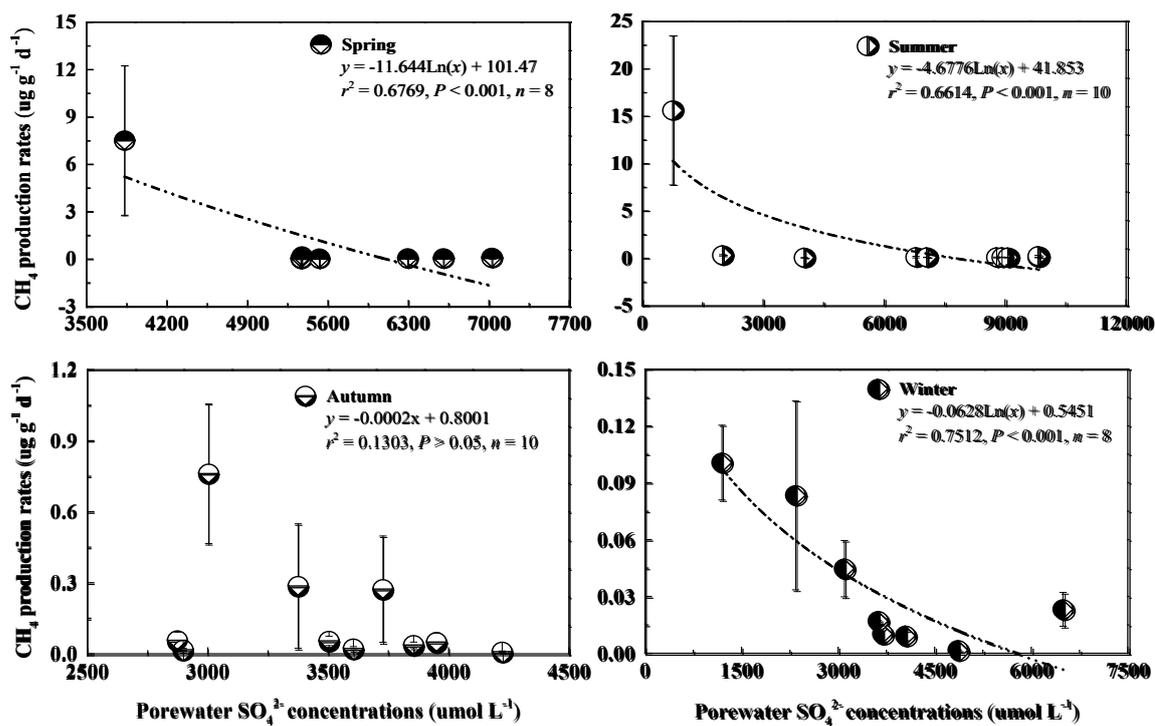


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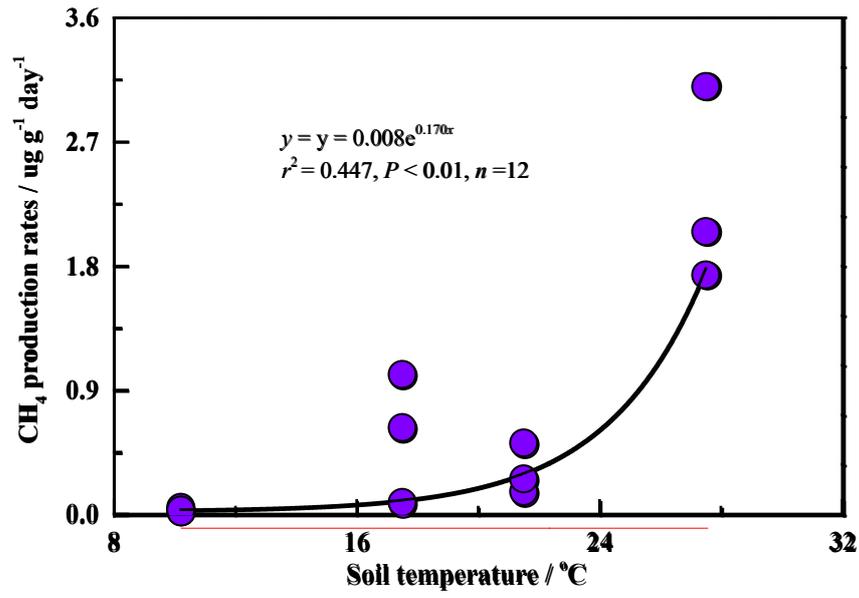


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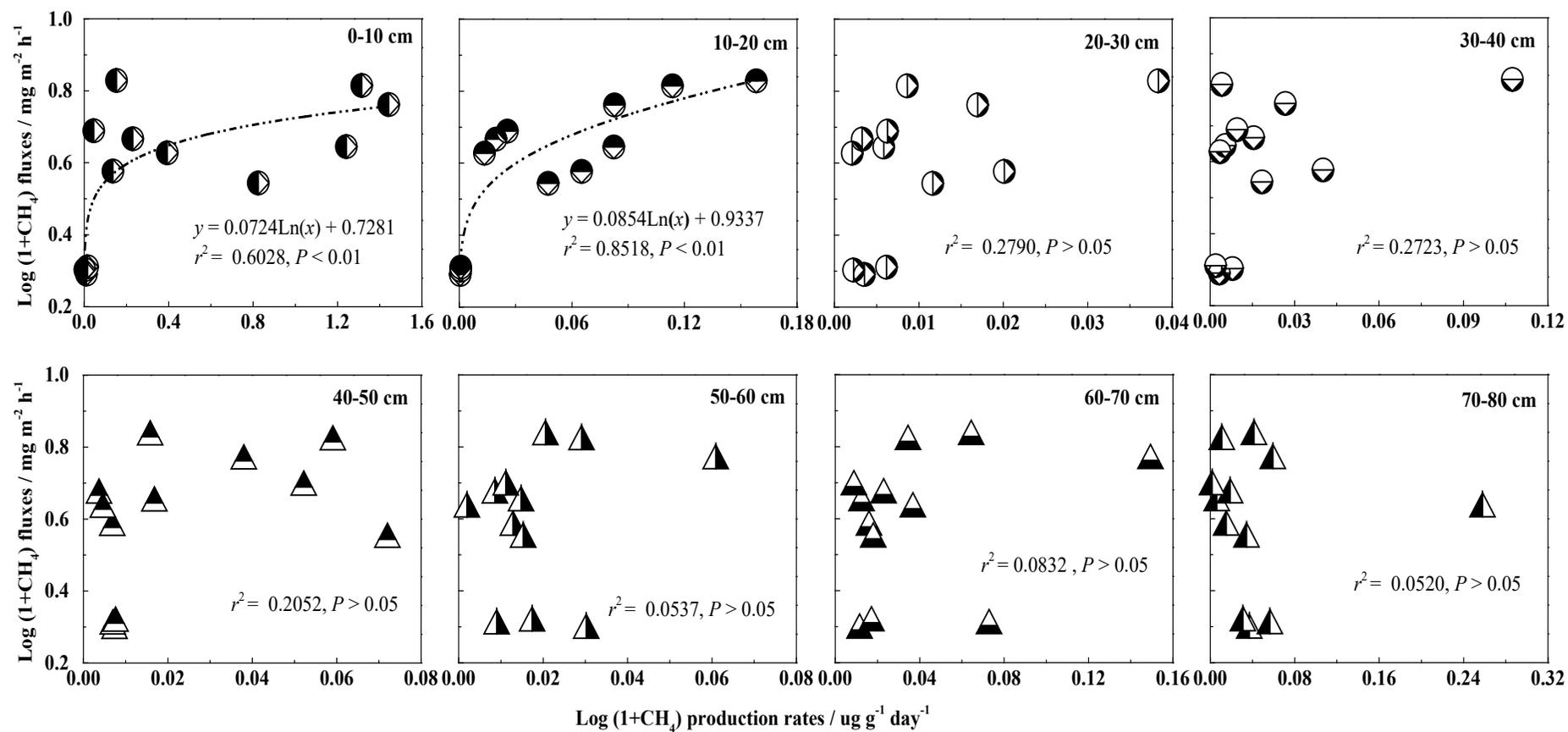


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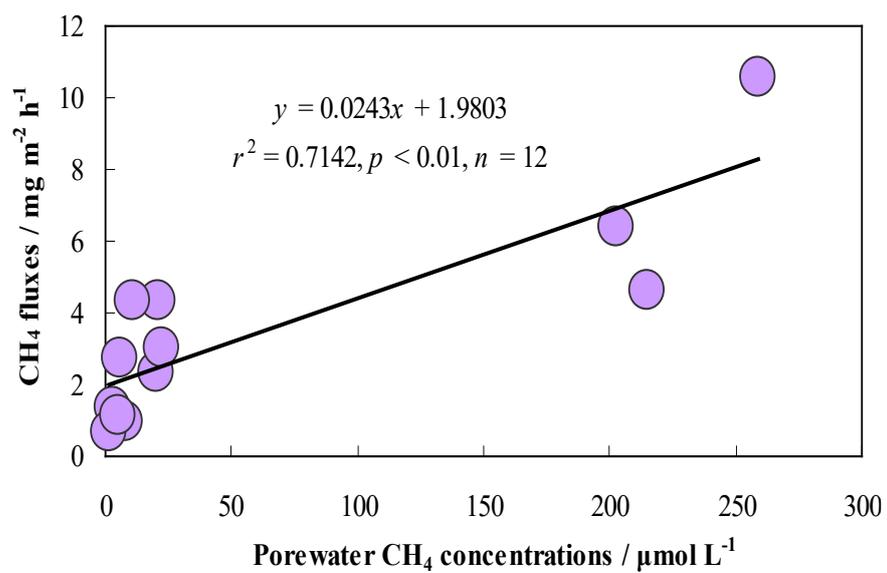


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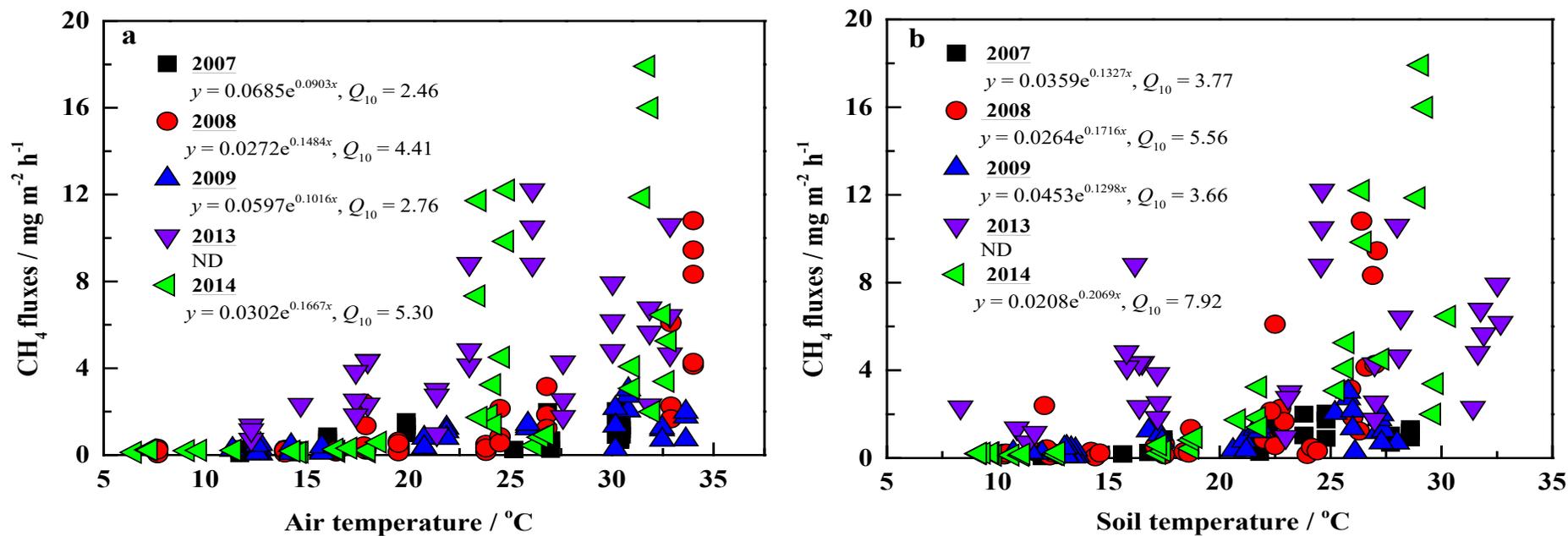


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